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  The number of right parentheses in a query must be equal to the number of left parentheses.
- => ("fluorescent resonance energy transfer" or FRET) and histone and modification
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  - 1 FILE BIOTECHABS
  - 1 FILE BIOTECHDS
  - 17 FILES SEARCHED...
    - 1 FILE CAPLUS
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  - 29 FILES SEARCHED...
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  - 60 FILES SEARCHED...
    - 2 FILE SCISEARCH
    - 263 FILE USPATFULL
    - 21 FILE USPAT2

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- L1 QUE ("FLUORESCENT RESONANCE ENERGY TRANSFER" OR FRET) AND HISTONE AND MODI FICATION

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=> ("fluorescent resonance energy transfer" or FRET) and histone and modification
L2 6 ("FLUORESCENT RESONANCE ENERGY TRANSFER" OR FRET) AND HISTONE
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=> d ti 1-6

- L3 ANSWER 1 OF 6 SCISEARCH COPYRIGHT (c) 2005 The Thomson Corporation on STN
- TI A fluorescence resonance energy transfer-based probe to monitor nucleosome structure
- L3 ANSWER 2 CF 6 CAPLUS COPYRIGHT 2005 ACS on STN

- TI Genetically encoded fusion protein fluorescent reporters of kinase, methyltransferase, and acetyltransferase activities in cells and tissues
- L3 ANSWER 3 OF 6 Elsevier BIOBASE COPYRIGHT 2005 Elsevier Science B.V. on STN
- TI A genetically encoded fluorescent reporter of **histone** phosphorylation in living cells
- L3 ANSWER 4 OF 6 Elsevier BIOBASE COPYRIGHT 2005 Elsevier Science B.V. on STN
- TI Modulation of DNA conformations through the formation of alternative high-order HU-DNA complexes
- L3 ANSWER 5 OF 6 SCISEARCH COPYRIGHT (c) 2005 The Thomson Corporation on STN
- TI Interaction of maize Opaque-2 and the transcriptional co-activators GCN5 and ADA2, in the modulation of transcriptional activity
- L3 ANSWER 6 OF 6 BIOSIS COPYRIGHT (c) 2005 The Thomson Corporation on STN
- TI Selective recognition of acetylated **histones** by bromodomain proteins visualized in living cells.

## => d ab bib 1-6

- L3 ANSWER 1 OF 6 SCISEARCH COPYRIGHT (c) 2005 The Thomson Corporation on STN
- AΒ Nucleosomes are the basic units of eukaryotic chromatin structure. By restricting factor access to regulatory DNA sequences, nucleosomes significantly impact genomic processes such as transcription, and various mechanisms to alter nucleosome structure to relieve this repression have evolved. Both nucleosomes and processes that alter them are inherently dynamic in nature. Thus, studies of dynamics will be necessary to truly. understand these relief mechanisms. We describe here the characteristics of a novel fluorescence resonance energy transfer-based reporter that can clearly signal the formation of a canonical nucleosome structure and follow conformational and compositional changes in that structure, both at the ensemble-average (bulk) and at the single molecule level. Labeled nucleosomes behave conformationally and thermodynamically like typical nucleosomes; thus they are relevant reporters of nucleosome behavior. Nucleosomes and free DNA are readily distinguishable at the single-molecule level. Thus, these labeled nucleosomes are well suited to studies of dynamic changes in nucleosome structure including single-molecule dynamics. © 2005 Elsevier Inc. All rights reserved.
- AN 2005:525011 SCISEARCH
- GA The Genuine Article (R) Number: 926KR
- TI A fluorescence resonance energy transfer-based probe to monitor nucleosome structure
- AU Lovullo D; Daniel D; Yodh J; Lohr D; Woodbury N W (Reprint)
- CS Arizona State Univ, Dept Chem & Biochem, Tempe, AZ 85287 USA (Reprint); Midwestern Univ, Coll Osteopath Med, Div Basic Sci, Glendale, AZ 85308 USA; Arizona State Univ, Biodesign Inst, Tempe, AZ 85287 USA nwoodbury@asu.edu
- CYA USA
- SO ANALYTICAL BIOCHEMISTRY, (1 JUN 2005) Vol. 341, No. 1, pp. 165-172. ISSN: 0003-2697.
- PB ACADEMIC PRESS INC ELSEVIER SCIENCE, 525 B ST, STE 1900, SAN DIEGO, CA 92101-4495 USA.
- DT Article; Journal
- LA English
- REC Reference Count: 53
- ED Entered STN: 2 Jun 2005
  - Last Updated on STN: 2 Jun 2005
  - \*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

- L3 ANSWER 2 OF 6 CAPLUS COPYRIGHT 2005 ACS on STN
- The invention provides fusion protein reporter mols. that can be used to AB monitor protein modifications (e.g., histone modifications) in living cells, and methods of using the fusion  ${\tt reporter\ mols.}\ for\ {\tt diagnosing\ protein-modification-associated}$ disorders (e.g. histone-modification-associated disorders). Reporters are designed by fusing, in order from N- to C-terminus, cyan fluorescent protein (CFP), a binding domain specific for the modified histone sequence of interest, a peptide substrate corresponding to the N-terminus of histone H3 or H4, and yellow fluorescent protein (YFP). Modification of the peptide substrate by a kinase, acetyltransferase, or methyltransferase then allows it to form an intramol. complex with the binding domain, increasing fluorescence resonance energy transfer (FRET) between the two flanking fluorescent moieties. Removal of the modification by a phosphatase, deacetylase, or (if methylation is reversible) demethylase reverses the FRET change. This design is optimized empirically to maximize responsivity by interchanging the donor and acceptor or the substrate and binding domain, or by varying the length and content of interdomain spacer sequences (linker sequences). Gcn5-based and TAFAB-based histone acetylation reporters are emphasized. invention also provides methods of using the fusion protein reporters to identify candidate pharmaceutical agents that effect protein modification in cells and tissues, thus permitting identification of candidate pharmaceutical agents for treatment of proteinmodification-associated disorders.
- AN 2004:430935 CAPLUS
- DN 141:18691
- TI Genetically encoded fusion protein fluorescent reporters of kinase, methyltransferase, and acetyltransferase activities in cells and tissues
- IN Ting, Alice Y.
- PA Massachusetts Institute of Technology, USA
- SO PCT Int. Appl., 96 pp. CODEN: PIXXD2
- DT Patent
- LA English
- FAN.CNT 1

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			ΙT,	LU,	MC,	NL,	PT,	RO,	SE,	SI,	SK,	TR						
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	US	2003	-634	740		Α		2003	0805									

- ANSWER 3 OF 6 Elsevier BIOBASE COPYRIGHT 2005 Elsevier Science B.V. on STN
- AB An increase in FRET indicates phosphorylation of histone H3 at serine 28. The protein-based reporter (see picture) responds to phosphorylation through intramolecular complexation between a substrate domain derived from histone H3 and a linked phosphoserine-recognition domain. The reporter is also effective inside living mammalian cells. FRET = fluorescence resonance energy transfer.
- AN 2004244172 ESBIOBASE
- TI A genetically encoded fluorescent reporter of **histone** phosphorylation in living cells
- AU Lin C.-W.; Ting A.Y.
- CS Prof. A.Y. Ting, Department of Chemistry, Massachusetts Inst. of

Technology, Cambridge, MA 02139, United States.

E-mail: ating@mit.edu

SO Angewandte Chemie - International Edition, (24 MAY 2004), 43/22 (2940-2943), 15 reference(s)

CODEN: ACIEAY ISSN: 1433-7851

- DT Journal; Article
- CY Germany, Federal Republic of
- LA English
- SL English
- L3 ANSWER 4 OF 6 Elsevier BIOBASE COPYRIGHT 2005 Elsevier Science B.V. on STN
- AB HU is an abundant, highly conserved protein associated with the bacterial chromosome. It belongs to a small class of proteins that includes the eukaryotic proteins TBP, SRY, HMG-I and LEF-I, which bind to DNA non-specifically at the minor groove. HU plays important roles as an accessory architectural factor in a variety of bacterial cellular processes such as DNA compaction, replication, transposition, recombination and gene regulation. In an attempt to unravel the role this protein plays in shaping nucleoid structure, we have carried out fluorescence resonance energy transfer measurements of HU-DNA oligonucleotide complexes, both at the ensemble and single-pair levels. Our results provide direct experimental evidence for concerted DNA bending by HU, and the abrogation of this effect at HU to DNA ratios above about one HU dimer per 10-12 bp. These findings support a model in which a number of HU molecules form an ordered helical scaffold with DNA lying in the periphery. The abrogation of these nucleosome-like structures for high HU to DNA ratios suggests a unique role for HU in the dynamic modulation of bacterial nucleoid structure. .COPYRGT. 2004 Elsevier Ltd. All rights reserved.
- AN 2004193691 ESBIOBASE
- TI Modulation of DNA conformations through the formation of alternative high-order HU-DNA complexes
- AU Sagi D.; Friedman N.; Vorgias C.; Oppenheim A.B.; Stavans J.
- CS J. Stavans, Dept. of Physics of Complex Systems, The Weizmann Institute of Science, Rehovot, Israel.
  E-mail: joel.stavans@weizmann.ac.il
- SO Journal of Molecular Biology, (06 AUG 2004), 341/2 (419-428), 41 reference(s)
  CODEN: JMOBAK ISSN: 0022-2836
- PUI S0022283604006916
- DT Journal; Article
- CY United Kingdom
- LA English
- SL English
- L3 ANSWER 5 OF 6 SCISEARCH COPYRIGHT (c) 2005 The Thomson Corporation on STN
- AB Maize Opaque-2 (ZmO2), a bZip class transcription factor has been shown to activate the transcription of a series of genes expressed in the maturation phase of endosperm development. Activation requires the presence of one or more enhancer binding sites, which confer the propensity for activation by ZmO2 on heterologous promoters and in heterologous plant cell types, such as tobacco mesophyll protoplasts. region of ZmO2 required for conferring transcriptional activation has been localised to a stretch of acidic residues in the N-terminal portion of the ZmO2 sequence, which is conserved between O2-related bZip factor sequences. Previously we identified the maize homologues of yeast transcriptional co-activators GCN5 and ADA2 that are implicated in nucleosome modification and transcription. In the present study we have shown that transcriptional modulation by ZmO2 involves the intranuclear interaction of ZmO2 with ZmADA2 and ZmGCN5. Forster resonance energy transfer (FRET) based techniques have enabled us to estimate the intracellular site of these intermolecular

interactions. As a functional readout of these intranuclear interactions, we used the ZmO2 responsive maize b-32 promoter to drive the beta-glucuronidase (GUS) in the presence and absence of ZmGCN5 and ZmADA2. Our results suggest that the likely recruitment of ZmADA2 and ZmGCN5 modulates the transactivation of b-32 promoter by ZmO2 and that there may be a competition between ZmGCN5 and ZmO2 for binding to the amino-terminal of ZmADA2. The results may be taken as a paradigm for other processes of transcriptional modulation in planta involving acidic activation domains.

- AN 2005:19126 SCISEARCH
- GA The Genuine Article (R) Number: 879AZ
- TI Interaction of maize Opaque-2 and the transcriptional co-activators GCN5 and ADA2, in the modulation of transcriptional activity
- AU Bhat R A; Borst J W; Riehl M; Thompson R D (Reprint)
- CS INRA, Res Unit Genet & Ecophysiol Grain Legumes URLEG, BP 86510, F-21065 Dijon, France (Reprint); Max Planck Inst Plant Breeding Res, D-50829 Cologne, Germany; Univ Wageningen & Res Ctr, Microspectrometry Ctr, NL-6703 HA Wageningen, Netherlands thompson@epoisses.inra.fr
- CYA France; Germany; Netherlands
- SO PLANT MOLECULAR BIOLOGY, (MAY 2004) Vol. 55, No. 2, pp. 239-252. ISSN: 0167-4412.
- PB KLUWER ACADEMIC PUBL, VAN GODEWIJCKSTRAAT 30, 3311 GZ DORDRECHT, NETHERLANDS.
- DT Article; Journal
- LA English

L3

- REC Reference Count: 65
- ED Entered STN: 13 Jan 2005
  - Last Updated on STN: 13 Jan 2005
  - \*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FCRMATS\*
- AB Acetylation and other modifications on histones
  comprise histone codes that govern transcriptional regulatory
  processes in chromatin. Yet little is known how different histone
  codes are translated and put into action. Using fluorescence resonance
  energy transfer, we show that bromodomain-containing proteins recognize
  different patterns of acetylated histones in intact nuclei of
  living cells. The bromodomain protein Brd2 selectively interacted with
  acetylated lysine 12 on histone H4, whereas TAFdblvert250 and

ANSWER 6 OF 6 BIOSIS COPYRIGHT (c) 2005 The Thomson Corporation on STN

PCAF recognized H3 and other acetylated histones, indicating fine specificity of histone recognition by different bromodomains. This hierarchy of interactions was also seen in direct peptide binding assays. Interaction with acetylated histone was essential for Brd2 to amplify transcription. Moreover association of Brd2, but not other bromodomain proteins, with acetylated chromatin

persisted on chromosomes during mitosis. Thus the recognition of **histone** acetylation code by bromodomains is selective, is involved in transcription, and potentially conveys transcriptional memory across cell divisions.

- AN 2004:149090 BIOSIS
- DN PREV200400152814
- TI Selective recognition of acetylated **histones** by bromodomain proteins visualized in living cells.
- AU Kanno, Tomohiko; Kanno, Yuka; Siegel, Richard M.; Jang, Moon Kyoo; Lenardo, Michael J.; Ozato, Keiko [Reprint Author]
- CS Laboratory of Molecular Growth Regulation, National Institute of Child Health and Human Development, National Institutes of Health, Bethesda, MD, 20892, USA ozatok@nih.gov
- SO Molecular Cell, (January 16 2004) Vol. 13, No. 1, pp. 33-43. print. ISSN: 1097-2765 (ISSN print).
- DT Article
- LA English
- ED Entered STN: 17 Mar 2004